LONG CHAIN ESTERS OF AESCULUS INDICA¹

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In continuation of our chemical investigation on Indian horse chestnuts, seeds of Aesculus indica L. (Hippocastanaceae) (1,2), we have now isolated two unsaturated and two saturated aliphatic esters, designated as compounds A, B, C, and D, from the CHCl₃ extract.

Compound A, a viscous liquid, had ir absorption bands at 2910, 2850, and 715 cm⁻¹ for its long chain, at 1735 cm⁻¹ for ester carbonyl, and at 1685 cm⁻¹ for an isolated double bond. Its ¹H nmr exhibited a pair of olefinic methine tion around 970 cm⁻¹ in the ir spectrum. An oxymethylene quartet (J = 6.0 Hz) at $\delta 4.04$ in addition to a broad triplet (J = 6.0 Hz) at $\delta 0.81$ due to methyl groups (one terminal) led us to predict the presence of an OCH₂CH₃ substituent as an esterifying residue (5).

Its mass spectrum had a $[M]^+$ at m/z310 revealing the molecular formula $C_{20}H_{38}O_2$. The ion at m/z 45 and the loss of 45 mass units from the $[M]^+$ resulting in the formation of an ion at m/z 265 were characteristic of a terminal

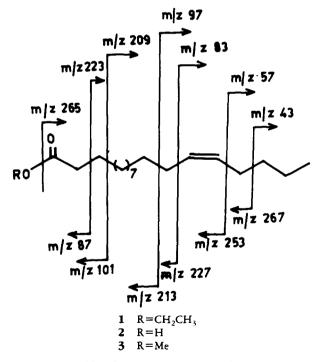


FIGURE 1. Mass Fragmentation Pattern of 1, 2, and 3.

signals at δ 5.25 as broad doublets (J = 11 Hz) of triplets (J = 5.5 Hz), hence revealing the *cis* geometry of the olefinic bond (3,4) which is moreover supported by the absence of an absorp-

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ethoxyl group (5). This conclusion was further substantiated by the presence of an ion at m/z 87 corresponding to $CH_2COOCH_2CH_3$ (5). The location of the double bond at C-13 was determined from prominent β -fission (6) ions obtained at m/z 57, 253, 83, and 227 and α -fission (6) ions at 43, 267, 97, and 213 (Figure 1). There were fragments corresponding to the sequential loss of 14 mass units between m/z 223 and m/z83 which revealed the existence of a straight chain up to C-12 followed by double bond at C-13.

On alkaline hydrolysis compound A yielded an acid 2, which showed a carboxylic group absorption band at 1710 cm^{-1} in its ir spectrum and the absence of a quartet at δ 4.04 and the presence of a sharp triplet at δ 0.85 in its ¹H-nmr spectrum. Its ms exhibited $[M]^+ m/z$ 282 corresponding to C18H34O2, fragments due to the loss of 45 and 60 mass units, and remaining fragments which were quite similar to compound A. This indicated a C18 straight chain with olefinic unsaturation at C-13 which was in accordance with the appearance of both olefinic methines at δ 5.30 ($J_{13,14} = 11$ Hz, $J_{12,13} = J_{14,15} = 5.5$ Hz). Thus, 2 could be identified as octadeca-(13Z)enoic acid and consequently compound

A as ethyl octadec-(13Z)-enoate [1], which was in full accordance with its ^{13}C shielding data.

The ¹³C-nmr spectrum of compound A (Table 1) displayed resonances at δ 173.42 for ester carbonyl, δ 129.73 and δ 129.51 for olefinic methines, δ 33.97 for a methylene adjacent to an ester carbonyl group, and δ 59.98 and δ 14.01 corresponding to an OCH2CH3 grouping. The definitive ¹³C signal assignments were made by comparison with the related n-alkane (7,8) and with other similar esters (9-11). The Z configuration of the olefinic bond (12-14) was indicated by the appearance of an allylic methylene at δ 27.01. The remaining thirteen methylene signals were observed between δ 31.71 and 24.76 together with the signals at δ 22.46 and 14.01 due to a terminal CH₂CH₃ group. This confirmed the presence of a C_{18} straight chain acid with an olefinic unsaturation at C-13 and an ethyl group as an ester moiety, thus unequivocally

Carbon .	Compound			
	1	3	ethyl heptadecanoate	methyl heptadecanoate
1	175.68	173.42	175.77	173.46
2	33.97	33.91	33.95	33.66
3	24.76	24.78	24.66	24.66
4	28.81	28.91	28.91	28.91
5	29.55 ^ª	29.49 ^a	29.83ª	29.69ª
6	29.47ª	29.49ª	29.03ª	29.03ª
7	29.32 ^ª	29.49ª	29.23°	29.23ª
8	29.11 ^a	29.14 ^a	29.44 ^a	29.44ª
9	29.06ª	29.14 ^a	29.44 ^a	29.44 ^a
10	28.94ª	28.96ª	29.44ª	29.44ª
11	29.11 ^a	28.96ª	29.44 ^a	29.44ª
12	27.01	27.02	29.44 ^a	29.44ª
13	129.73	129.79	29.12 ^a	29.12 ^a
14	129.51	129.54	29.91ª	29.91*
15	27.01	27.02	31.68	31.68
16	31.71	31.73	22.39	22.39
17	22.46	22.48	12.68	13.68
18	13.87	13.87	_	— —
ОСН,СН,	59.98	—	59.58	
$\mathbf{CH}_{\mathbf{CH}_{\mathbf{T}}}$	14.01	—	13.86	_
ОМе ⁻	_	51.15	_	50.76

 TABLE 1.
 13C-nmr Data for Ethyl Octadec-(13Z)-enoate [1], Methyl octadec-(13Z)-enoate [3], Ethyl Heptadecanoate, and Methyl Heptadecanoate.

^aAssignments may be reversed in any vertical column.

supporting the proposed structure 1 for Per

compound A. Compound B, $C_{19}H_{36}O_2$ ([M]⁺ m/z296), exhibited a close resemblance of its ir, ¹H-, and ¹³C-nmr spectral data with those for compound A. However, instead of ethoxymethylene at δ 4.04, it showed a sharp singlet at δ 3.62 due to carboxymethyl group in its ¹H-nmr spectrum; we therefore inferred that it was a methyl ester. The presence of a strong peak at m/z 74 arose by McLafferty rearrangement, and a fragment ion at m/z 222, due to the rest of the molecule. moreover confirmed its methyl ester behavior. This was further supported by the presence of the signals at δ 175.68 and δ 51.15 assignable to a carboxymethyl group in its ¹³C-nmr spectrum, whereas remaining ¹³C spectral data (Table 1) was quite similar to that of compound A. Alkaline hydrolysis yielded an acid identified as 2. Hence, compound B can be identified as methyl octadec-(13Z)-enoate [3].

Compound C, a colorless viscous liquid, analyzed for $C_{19}H_{38}O_2$ and showed ir absorption bands at 1732 and 1270 cm⁻¹ for the ester group. The presence of a quartet at δ 4.01 and a broad triplet at δ 0.80 suggested it to be an ethyl ester. This was supported by its ms, which exhibited [M]⁺ at m/z 298 and fragment ions due to the loss of m/z 45 and m/z 73 and other fragments consistent with its identity as ethyl heptadecanoate.

Compound D, $C_{18}H_{34}O_2$ ([M]⁺ m/z 284), exhibited significant resemblance of its ir, ¹H-, and ¹³C-nmr spectral data with those for compound C except for the replacement of an ethoxyl group by a methoxyl group (¹H nmr δ 3.55, 3H; ¹³C nmr, δ 50.76). Based on its ms and ¹³C-nmr spectral data, it was identified as methyl heptadecanoate.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.— Melting points were taken on a Toshniwal melting point apparatus and are not corrected. Spectra were recorded with the following instruments: ir, Perkin-Elmer 399B model; ¹H nmr and ¹³C nmr, Varian Ft-80A in CDCl₃ with TMS as an internal standard. Si gel (60–120 mesh, BDH) was used for cc. All R_f values were reported on tlc using Si gel G. Plates (0.075 mm) were used for preparative layer chromatography.

PLANT MATERIAL AND EXTRACTION.-The seeds of A. indica were collected from Kulu, Himachal Pradesh, India, during the autumn of 1983 and identified at the Botany Department of this Institute. A voucher specimen is deposited in CIMAP herbarium collection, No. 1851. The air-dried, powdered seeds (1 kg) were defatted with *n*-hexane (5×2.5 liters) and then extracted with CHCl₃ $(5 \times 3$ liters). The total CHCl₃ extract was concentrated under reduced pressure to give a brown viscous mass (25 g), which was chromatographed over Si gel (500 g) and eluted with CHCl₃ and an increasing amount of MeOH in CHCl₃. The fractions 2-4 obtained by elution with CHCl₃ were combined and subjected to repeated cc followed by tlc [petroleum ether-C6H6 (8:2)} to yield compound A (105 mg), compound B (55 mg), compound C (85 mg), and compound D (42 mg).

Compound A [1].—Colorless viscous liquid, ir ν max (neat) 2930, 2855, 1735, 1460, 1440, 1365, 1240, 1170, 720 cm⁻¹; ¹H nmr δ 0.81 (6H, t, J = 6 Hz, OCH₂CH₃, H₃-18), 1.19 (24H, br s, 12×CH₂), 1.95 (4H, m, H₂-12, 15), 2.21 (2H, t, J = 6 Hz, H₂-2), 4.04 (2H, q, J = 6Hz, OCH₂CH₃), 5.25 (2H, dt, $J_{13,14} = 11$ Hz, $J_{12,13} = J_{14,15} = 5.5$ Hz, H-13, 14); ms m/z 310, 296, 284, 267, 264, 255, 253, 239, 227, 222, 213, 199, 185, 171, 157, 143, 129, 115, 100, 97, 87, 83, 69, 55, 43, 41; ¹³C nmr see Table 1.

Alkaline bydrolysis of compound A.—Compound A [1] (50 mg) was refluxed with 5% aqueous KOH solution, worked up as usual, and on crystallization from a CHCl₃/MeOH mixture yielded colorless crystals of the acid 2: mp 56°; ir ν max (KBr), 2920, 2840, 1710, 1460, 1410, 720 cm⁻¹; ¹H nmr δ 0.85 (3H, t, J = 6 Hz, H₃-18), 1.24 (24H, br s, 12 × CH₂), 1.98 (4H, m, H₂-12, 15), 2.31 (2H, t, J = 6 Hz, H₂-2), 5.30 (2H, t, J = 6 Hz, H-13, 14); ms m/z 282, 264, 256, 227, 222, 213, 219, 185, 171, 157, 143, 129, 115, 111, 101, 97, 86, 82, 73, 68, 57, 55, 43, 41.

Compound B [3].—Colorless liquid, ir ν max (near) 2930, 2855, 1740, 1460, 1435, 1360, 1250, 1170, 720 cm⁻¹; ¹H nmr δ 0.86 (3H, t, J = 6 Hz, H₃-18), 1.12 (24H, br s, 12 × CH₂), 1.98 (4H, m, H₂-12,15), 2.26 (2H, t, J = 6 Hz, H₂-2), 3.62 (3H, s, COOMe), 5.28 (2H, t, J = 5.5 Hz, 6 Hz, H-13,-14); ms m/z 296, 271, 265, 239, 228, 223, 213, 200, 199, 185, 171, 157, 143, 129, 115, 111, 101, 97, 87, 83, 74, 69, 57, 55, 43, 41; ¹³C nmr see Table 1. Alkaline bydrolysis of compound B.—Compound B (20 mg) was hydrolyzed with 5% aqueous KOH solution and worked up as usual. The acid was identified as 2 (mp, co-tlc, ir, ¹H nmr).

Compound C (etbyl beptadecanoate).—Colorless liquid; ir $\nu \max$ (neat) 2920, 2825, 1732, 1440, 1350, 1240, 1140, 670 cm⁻¹; ¹H nmr δ 0.80 (6H, t, J = 6 Hz, CH₂CH₃, OCH₂CH₃), 1.58 (28H, br s, 14×CH₂), 2.19 (2H, t, J = 6 Hz, OCOCH₂CH₂), 4.01 (2H, q, J = 6 Hz, OCOCH₂CH₃); ms m/z 298, 271, 256, 242, 226, 228, 212, 198, 184, 170, 156, 142, 128, 114, 100, 86, 74, 57, 55, 43; ¹³C nmr see Table 1.

Compound D (methyl beptadecanoate).—Colorless liquid, ir ν max (neat) 2920, 2848, 1730, 1440, 1410, 1352, 1240, 1145, 670 cm⁻¹; ¹H nmr δ 0.80 (3H, t, J = 6 Hz, CH₂CH₃), 1.58 (28H, br s, 14 × CH₂), 2.19 (2H, t, J = 6 Hz, OCOCH₂CH₃), 3.55 (3H, s, COOCH₃); ms m/z 284, 226, 212, 198, 184, 156, 142, 128, 100, 86, 74, 59, 43; ¹³C nmr see Table 1.

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